

# Synthesis of 16,18-Dihydroxycleroda-3,13Z-dien-16,15-olide, (+)-16-Hydroxycleroda-3,13Z-dien-16,15-olide, and (–)-Hydroxyhalima-5(10),13-dien-16,15-olide from (+)-Hardwickiic Acid

Paulo M. Imamura<sup>\*,†</sup> and Marta Costa,<sup>‡,1</sup>

Instituto de Química, Universidade Estadual de Campinas, C.P. 6154, CEP 13083-970, Campinas, São Paulo, Brazil, and Departamento de Química, UFMS, MS, Brazil

Received February 29, 2000

Syntheses of three enantiomers of natural hydroxybutenolide diterpenes, 16,18-dihydroxycleroda-3,13Z-dien-16,15-olide (**4**), (+)-16-hydroxycleroda-3,13Z-dien-16,15-olide (**5**), and (–)-16-hydroxyhalima-5(10),-13Z-dien-16,15-olide (**6**), via a furan photosensitized oxygenation reaction of (+)-hardwickiic acid (**2**), are described.

The occurrence of natural clerodane hydroxybutenolides such as **1** (Figure 1) has been reported frequently in plants of the genera *Polyalthia*,<sup>2–11</sup> *Acritopappus*,<sup>12</sup> *Premna*,<sup>13</sup> and *Cyathocalyx*.<sup>14</sup> It is interesting to note that many species of these genera are widely used in folk medicine as diuretics,<sup>15</sup> febrifuges,<sup>4</sup> and chewing sticks, and for sterilizing milk containers.<sup>13</sup> It is also noteworthy that some natural clerodane hydroxybutenolides show significant biological activities as antifeedants<sup>2</sup> and antimicrobials,<sup>6,13</sup> cytotoxicity to tumor cell cultures,<sup>3,10</sup> and toxicity against *Artemia salina*<sup>3,14</sup> and *Aedes aegypti*.<sup>14</sup> Recently, we described the isolation of two clerodane hydroxybutenolides from *Echinodorus grandiflorus* [Cham. & Schldt.] Micheli (*Alismataceae*) and the synthesis of an enantiomer of one of them from (+)-hardwickiic acid (**2a**).<sup>15</sup> Although there are many reports in the literature for the synthesis of clerodane diterpenoids, only a few report the synthesis of clerodane hydroxybutenolides such as **1**<sup>16,17</sup> and **3**.<sup>15</sup> Thus, in connection with our studies regarding the use of resin acids as chiral starting materials for the synthesis of natural products,<sup>18–20</sup> and in view of the potential biological activities of natural hydroxybutenolides, we have synthesized clerodanes **4** and **5** and halimane **6**, with absolute configurations enantiomeric<sup>21</sup> to those of the natural products, from the readily available methyl (+)-hardwickiate (**2b**),<sup>15,22</sup> in order to facilitate comparisons of biological activities.

## Results and Discussion

Reduction of (+)-methyl hardwickiate (**2b**) with lithium aluminum hydride furnished a mixture (3:1) of the desired compound **7** and the corresponding  $\Delta^3$ -dihydro compound, which are difficult to separate. However, reduction of **2b** with diisobutylaluminum hydride (DIBAL) furnished the desired alcohol **7** in 98% yield (Scheme 1). The next step for the synthesis of the hydroxybutenolide moiety from the furan ring was based on a photosensitized oxygenation procedure reported by Kernan and Faulkner.<sup>23</sup> Thus, reaction of **7** in  $\text{CH}_2\text{Cl}_2$  at  $-78^\circ\text{C}$  with oxygen in the presence of Rose Bengal and diisopropylethylamine (DIPEA) furnished **4**, a diastereoisomer of a compound reported by Faulkner,<sup>23</sup> in 75% yield.

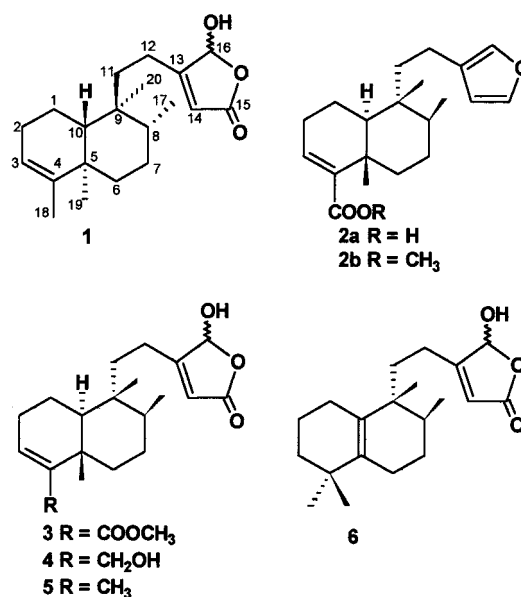
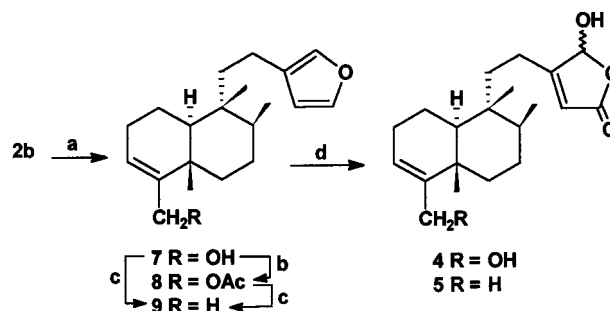


Figure 1.

## Scheme 1<sup>a</sup>



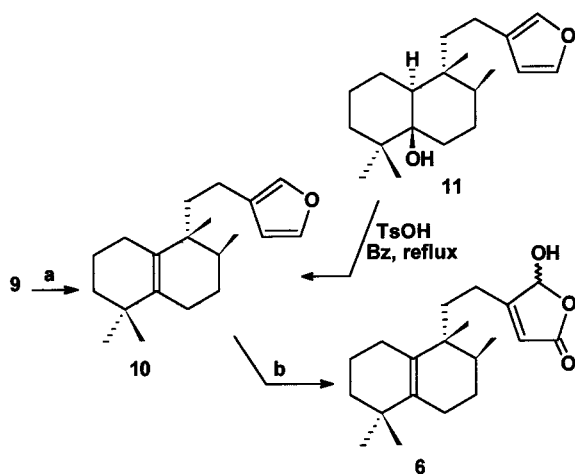
<sup>a</sup> Key: (a) DIBAL, toluene,  $-78^\circ\text{C}$  (98%); (b)  $\text{Ac}_2\text{O}$ , Py, rt, (85%); (c) nickel-boride, diglyme,  $0^\circ\text{C}$  (73% from **7** and 75% from **8**); (d)  $\text{O}_2$ , Rose Bengal, DIPEA,  $\text{CH}_2\text{Cl}_2$ ,  $-78^\circ\text{C}$ , (75% for **4** and 72% for **5**).

Reductive elimination of the hydroxyl group in **7** to yield **9** proved troublesome. Treatment of **7** with methanesulfonyl chloride led to a complex mixture of products, among which the desired mesylate and the corresponding chlorinated compound were tentatively identified. To circumvent this problem, Sharma's protocol<sup>24</sup> of transforming an allylic alcohol directly to the corresponding hydrocarbon was applied. Thus, treatment of **7** with nickel boride, generated

\* To whom correspondence should be addressed. E-mail: imam@iqm.unicamp.br.

<sup>†</sup> Universidade Estadual de Campinas.

<sup>‡</sup> Universidade Federal de Mato Grosso do Sul (UFMS).

Scheme 2<sup>a</sup>

<sup>a</sup> Key: (a) HCl–HOAc (1:4), 60 °C (85%); (b) O<sub>2</sub>, Rose Bengal, DIPEA, CH<sub>2</sub>Cl<sub>2</sub>, –78 °C, (90%).

in situ, gave **9** as a single product in 73% yield. Treatment of the acetate **8** with nickel boride also led to the desired compound **9** in 75% yield. Following the same procedure described above for **7**, photosensitized oxygenation of **9** gave **5** in 72% yield. Physical and spectroscopic data of the synthetic product were in good agreement with those reported for the natural product,<sup>2–13</sup> except for the expected difference in the optical rotation, which was  $[\alpha]_D +30.0$  (*c* 1.7, CHCl<sub>3</sub>) {lit.<sup>7</sup>  $[\alpha]_D -48.7^\circ$  (*c* 3.02, CHCl<sub>3</sub>)}.

Compound **10**, required for the synthesis of **6**, was obtained in 85% yield through acid-catalyzed rearrangement of **9** using acetic acid and hydrochloric acid at 60 °C (Scheme 2). Spectroscopic data of **10** were in good agreement with those reported in the literature for dehydroambliol-B, a dehydration product obtained from the marine diterpene ambliol-B (**11**) by treatment with p-TsOH in benzene.<sup>25,26</sup>

Finally, photosensitized oxygenation of **10** furnished the desired compound **6** in 90% yield. Spectroscopic data of **6** were also in good agreement with those reported in the literature for the enantiomer, except for the expected difference in the optical rotation, which was  $[\alpha]_D -35.4^\circ$  (*c* 0.24, CHCl<sub>3</sub>) {lit.<sup>5</sup>  $[\alpha]_D +21.0^\circ$  (*c* 0.54, CHCl<sub>3</sub>)}.

Compounds **3**, **5**, and **6** were evaluated for antitubercular activity. Only **5** showed significant activity according to the TAACF protocol (12.5 μg mL<sup>-1</sup>), with 85% inhibition of *Mycobacterium tuberculosis*.<sup>27</sup>

## Experimental Section

**General Experimental Procedures.** <sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded in CDCl<sub>3</sub> solution at 300 and 75 MHz, respectively, with a Bruker AC 300/P spectrometer (internal standard TMS). IR spectra were recorded on a Perkin-Elmer 1600 series FT IR. MS spectra were obtained at 70 eV on an HP-5990/5970 system equipped with a J&W Scientific DB-5 fused silica capillary column (30 m × 0.25 mm × 0.25 μm). Elemental analyses were performed with a Perkin-Elmer CHN analyzer. Optical rotations were measured with a Carl Zeiss photoelectric polarimeter.

{(4*ac*,6*β*,8*αβ*)-1-Hydroxymethyl-5(*R*)-[2-(3-furanyl)ethyl]-5,6,8*a*-trimethyl-3,4,4*a*,5,6,7,8,8*a*-octahydronaphthalene} (**7**). A 1.0 M solution of DIBAL (3.7 mL, 3.7 mmol) was added to a stirred solution of **2b** (558 mg, 1.7 mmol) in dry toluene (20 mL), at –78 °C, and the reaction mixture was warmed to 0 °C and stirred for 2 h. Saturated aqueous ammonium chloride (8 mL) was added, and the reaction mixture was extracted with ethyl ether (3 × 30 mL), washed with brine, and dried over anhydrous magnesium sulfate. The solvent was

removed under reduced pressure, and the residue was chromatographed on silica gel (petroleum ether–ethyl acetate; 9:1) to give **7** (500 mg; 98%) as a colorless oil:  $[\alpha]^{23}_D +18.3$  (CHCl<sub>3</sub>, *c* 1.49); IR (neat)  $\nu_{\max}$  3422, 2926, 2867, 1654, 1639, 1458, 1383, 1264, 1024, 873, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.75 (3H, s, H-20), 0.83 (3H, d, *J* = 6.4 Hz, H-17), 1.08 (3H, s, H-19), 1.20–1.80 (11H, m), 2.00–2.40 (4H, m), 4.10 (2H, d, *J* = 1.5 Hz, H-18), 5.57 (1H, t, *J* = 1.7 Hz, H-3), 6.26 (1H, t, *J* = 1.0 Hz, H-14), 7.20 (1H, bs, H-16), 7.35 (1H, t, *J* = 1.7 Hz, H-15); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  148.2 (s, C-4), 142.9 (d, C-15), 138.6 (d, C-16), 125.9 (s, C-13), 122.3 (d, C-3), 111.2 (d, C-14), 63.1 (t, C-18), 46.4 (d, C-10), 38.8 (s, C-9), 38.6 (t, C-11), 37.5 (s, C-5), 36.4 (t, C-6), 36.3 (d, C-8), 27.3 (t, C-7), 26.6 (t, C-2), 21.4 (q, C-19), 18.3 (q, C-20), 18.2 (t, C-1), 18.2 (t, C-12), 16.0 (q, C-17); MS (*m/z*, %) (M<sup>+</sup>) 302(16), 271(16), 189(50), 94(58), 81(100), 41(70); *anal.* C 79.29%, H 9.74%, calcd for C<sub>20</sub>H<sub>30</sub>O<sub>2</sub>, C 79.42%, H 9.99%.

{(4*ac*,6*β*,8*αβ*)-1-Acetoxymethyl-5(*R*)-[2-(3-furanyl)ethyl]-5,6,8*a*-trimethyl-3,4,4*a*,5,6,7,8,8*a*-octahydronaphthalene} (**8**). Acetic anhydride (0.1 mL, 0.6 mmol) was added to a solution of **7** (75 mg, 0.2 mmol) in dry pyridine (1 mL), and the mixture was stirred overnight at room temperature. Cold water (10 mL) was added, and the reaction mixture was extracted with ethyl ether (3 × 30 mL). The organic phase was washed with 5% HCl (2 × 10 mL) and then with brine until pH 7.0 and dried over anhydrous sodium sulfate. After removal of solvent under reduced pressure and purification of the residue by column chromatography (silica gel, *n*-hexane), compound **8** (72 mg, 85%) was obtained as a colorless oil.  $[\alpha]^{23}_D +26.3$  (CHCl<sub>3</sub>, *c* 1.94); IR (neat)  $\nu_{\max}$  2925, 2856, 1740, 1450, 1384, 1239, 1026, 873, 780; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.75 (3H, s, H-20), 0.83 (3H, d, *J* = 6.5 Hz, H-17), 1.08 (3H, s, H-19), 1.30–1.80 (10H, m), 1.98–2.40 (4H, m), 2.07 (3H, s, OAc), 4.53 (2H, t, *J* = 2.7 Hz, H-18), 5.60 (1H, t, *J* = 3.5 Hz, H-3), 6.26 (1H, d, *J* = 1.8 Hz, H-16), 7.20 (1H, bs, H-14), 7.34 (1H, t, *J* = 1.6 Hz, H-15); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  171.3 (s, C=O), 142.9 (s, C-4), 142.9 (d, C-15), 138.6 (d, C-16), 126.2 (d, C-3), 125.9 (s, C-13), 111.2 (d, C-14), 65.1 (t, C-18), 46.3 (d, C-10), 38.8 (s, C-9), 38.6 (t, C-11), 37.9 (s, C-5), 36.3 (d, C-8), 36.2 (t, C-6), 27.2 (t, C-7), 26.7 (t, C-2), 21.3 (q, C-19), 21.2 (q, H<sub>3</sub>C–CO), 18.2 (t, C-1), 18.2 (t, C-12), 18.1 (q, C-20), 16.0 (q, C-17); *anal.* C 76.56%, H 9.14%, calcd for C<sub>22</sub>H<sub>32</sub>O<sub>3</sub>, C 76.70%, H 9.36%.

{(4*ac*,6*β*,8*αβ*)-5(*R*)-[2-(3-Furanyl)ethyl]-1,5,6,8*a*-tetramethyl-3,4,4*a*,5,6,7,8,8*a*-octahydronaphthalene} (**9**). Anhydrous nickel chloride (622 mg, 4.8 mmol) and sodium borohydride (363 mg, 9.6 mmol) were added to a solution of **7** (104 mg, 0.3 mmol) in diglyme (15 mL) at 0 °C, whereupon a black precipitate formed. The mixture was stirred for 6 h, then diluted with dichloromethane (20 mL), and filtered through a pad of Celite, and the solvent was removed under reduced pressure. The residue was chromatographed on silica gel (*n*-hexane–ethyl ether; 95:5) to give **9** (72 mg, 73%) as a colorless oil.  $[\alpha]^{23}_D +21.3$  (CHCl<sub>3</sub>, *c* 2.64); IR (neat)  $\nu_{\max}$  2925, 2866, 1458, 1382, 1160, 1025, 872, 778 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.73 (3H, s, H-20), 0.83 (3H, d, *J* = 6.3 Hz, H-17), 0.99 (3H, s, H-19), 1.10–1.90 (10H, m), 1.56 (3H, d, *J* = 1.6 Hz, H-18), 2.00–2.40 (4H, m), 5.11 (1H, bs, H-3), 6.14 (1H, s, H-16), 7.10 (1H, bs, H-14), 7.24 (1H, t, *J* = 1.5 Hz, H-15); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  143.7 (s, C-4), 142.3 (d, C-15), 138.0 (d, C-16), 125.2 (s, C-13), 120.6 (d, C-3), 110.7 (d, C-14), 46.1 (d, C-10), 38.5 (t, C-11), 38.4 (s, C-9), 37.9 (s, C-5), 36.6 (t, C-6), 36.2 (d, C-8), 27.4 (t, C-7), 26.7 (t, C-2), 19.8 (q, C-19), 18.2 (t, C-1), 18.2 (t, C-12), 18.2 (q, C-20), 17.9 (q, C-18), 16.1 (q, C-17); MS (*m/z*, %) (M<sup>+</sup>) 286(30), 271(35), 191(60), 107(62), 95(100), 81(80), 41(70); *anal.* C 83.57%, H 10.73%, calcd for C<sub>20</sub>H<sub>30</sub>O, C 83.86%, H 10.56%.

{(4*ac*,6*β*,8*αβ*)-1-Hydroxymethyl-5(*R*)-[2-(2,5-dihydro-5-hydroxy-2-oxo-4-furanyl)ethyl]-5,6,8*a*-trimethyl-3,4,4*a*,5,6,7,8,8*a*-octahydronaphthalene} (**4**). Oxygen was bubbled through a solution of **7** (80 mg, 0.3 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (20 mL), diisopropylamine (0.4 mL), and Rose Bengal on polystyrene (3 mg) and irradiated with a tungsten lamp (250 W) at –78 °C for 6 h. The reaction mixture was filtered through a pad of Celite, and the residue was purified by silica

gel column chromatography (CHCl<sub>3</sub>-MeOH; 99:1) to give **4** (66 mg, 75%) as an oil: IR (neat)  $\nu_{\max}$  3372, 2926, 1756, 1648, 1458, 1129, 737 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.78 (3H, s, H-20), 0.81 (3H, d,  $J$  = 6.0 Hz, H-17), 1.08 (3H, s, H-19), 1.20–2.50 (15H, m), 4.10 (2H, s, H-18), 4.55 (1H, br, OH), 5.57 (1H, s, H-3), 5.84 (1H, s, H-14), 6.00 (1H, s, H-16); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  171.5 (s, C-15), 170.4 (s, C-13), 142.3 (s, C-4), 136.8 (d, C-3), 117.0 (d, C-14), 99.1 (d, C-16), 63.1 (t, C-18), 46.5 (d, C-10), 38.7 (s, C-9), 37.5 (s, C-5), 36.3 (d, C-8), 35.7 (t, C-6), 34.8 (t, C-11), 27.1 (t, C-7), 27.0 (t, C-2), 21.2 (t, C-12), 20.6 (q, C-19), 18.1 (q, C-20), 17.4 (q, C-1), 15.9 (C-17); *anal.* C 71.59%, H 9.14%, calcd for C<sub>20</sub>H<sub>30</sub>O<sub>4</sub>, C 71.82%, H, 9.04%.

{(4 $\alpha$ ,6 $\beta$ ,8 $\alpha\beta$ )-5(R)-[2-(2,5-Dihydro-5-hydroxy-2-oxo-4-furanyl)ethyl]-1,5,6,8a-tetramethyl-3,4,4a,5,6,7,8,8a-octahydronaphthalene} (**5**). Using the same procedure as described above for **7**, reaction of **9** (30 mg, 0.1 mmol) furnished hydroxybutenolide **5** (24 mg, 72%) as an oil: [ $\alpha$ ]<sub>D</sub><sup>23</sup> +30.0 (c 1.7, CHCl<sub>3</sub>); IR (neat)  $\nu_{\max}$  3336, 2956, 1759, 1737, 1647, 1448, 1130, 953, 738 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.72–0.98 (2H, m), 0.77 (3H, s, H-20), 0.81 (3H, d,  $J$  = 6.0 Hz, H-17), 1.01 (3H, s, H-19), 1.10–1.84 (9H, m), 1.58 (3H, d,  $J$  = 1.5 Hz, H-18), 1.90–2.40 (3H, m), 4.58 (1H, br, OH), 5.19 (1H, s, H-3), 5.84 (1H, s, H-14), 6.01 (1H, s, H-16); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  171.6 (s, C-15), 170.4 (s, C-13), 144.4 (s, C-4), 122.4 (d, C-3), 117.0 (d, C-14), 99.1 (d, C-16), 46.5 (d, C-10), 38.7 (s, C-9), 38.7 (s, C-5), 38.2 (t, C-6), 36.7 (d, C-8), 34.8 (t, C-11), 27.4 (t, C-7), 26.5 (t, C-2), 21.4 (t, C-12), 19.9 (q, C-19), 18.3 (t, C-1), 18.2 (q, C-17), 18.0 (q, C-18), 16.0 (q, C-20); MS (*m/z*, %) (M<sup>+</sup>) 318(25), 275(70), 191(80), 149(70), 123(100), 107(98), 41(96); *anal.* C 75.29%, H 9.44%, calcd for C<sub>20</sub>H<sub>30</sub>O<sub>3</sub>, C 75.43%, H, 9.50%.

(6 $\beta$ )-5(R)-[2-(3-Furanyl)ethyl]-1,1,5,6-tetramethyl-1,2,3,4,5,6,7,8-octahydronaphthalene} (**10**). A solution of **9** (35 mg, 0.1 mmol) in acetic acid (0.4 mL) and concentrated hydrochloric acid (0.1 mL) was heated at 60 °C. After stirring for 12 h, water (5 mL) was added and the mixture was extracted with ethyl acetate (3 × 30 mL). The organic layer was washed with brine and dried over anhydrous sodium sulfate. After removal of solvent under reduced pressure and purification of the residue by column chromatography (silica gel, *n*-hexane), compound **10** (30 mg, 85%) was obtained as a colorless oil: [ $\alpha$ ]<sub>D</sub><sup>23</sup> -26.0 (CHCl<sub>3</sub>, c 1.47); IR (film) 2956, 1495, 1452, 1363, 1028, 730 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.83 (3H, s, H-20), 0.87 (3H, d,  $J$  = 6.9 Hz, H-17), 0.98 (3H, s, H-18), 0.99 (3H, s, H-19), 1.10–1.90 (11H, m), 2.00–2.40 (4H, m), 6.13 (1H, bs, H-14), 7.09 (1H, bs, H-16), 7.23 (1H, bs, H-15); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  142.9 (d, C-15), 138.8 (s, C-5), 137.5 (d, C-16), 132.8 (s, C-10), 126.1 (s, C-13), 111.3 (d, C-14), 40.9 (s, C-9), 40.3 (t, C-3), 36.8 (t, C-11), 34.7 (s, C-4), 33.8 (d, C-8), 29.4 (q, C-18), 27.8 (q, C-19), 27.5 (t, C-7), 26.2 (t, C-6), 25.6 (t, C-1), 21.2 (q, C-20), 20.3 (t, C-12), 19.9 (t, C-2), 16.3 (q, C-17); *anal.* C 83.69%, H 10.34%, calcd for C<sub>20</sub>H<sub>30</sub>O, C 83.86%, H 10.56%.

{(6 $\beta$ )-5(R)-[2-(2,5-Dihydro-5-hydroxy-2-oxo-4-furanyl)ethyl]-1,1,5,6-tetramethyl-1,2,3,4,5,6,7,8-octahydronaphthalene} (**6**). Using the same procedure as described for **7**, reaction of **10** (30 mg, 0.1 mmol) furnished hydroxybutenolide **6** (30 mg, 90%) as an oil: [ $\alpha$ ]<sub>D</sub><sup>23</sup> -35.4 (CHCl<sub>3</sub>, c 0.24); IR (film) 3386, 2944, 1753, 1648, 1256, 1128, 738 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  0.86 (3H, d,  $J$  = 6.6 Hz, H-17), 0.87 (3H, s, H-20),

0.97 (3H, s, H-18), 0.99 (3H, s, H-19), 1.10–2.50 (15H, m), 4.25 (1H, bs, OH), 5.84 (1H, s, H-14), 6.00 (1H, bs, H-16); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz)  $\delta$  171.3 (s, C-15), 170.6 (s, C-13), 138.5 (s, C-5), 131.3 (s, C-10), 117.0 (d, C-14), 98.9 (d, C-16), 40.7 (s, C-9), 39.8 (t, C-3), 34.6 (s, C-4), 33.7 (d, C-8), 32.8 (t, C-11), 29.2 (q, C-18), 27.6 (q, C-19), 27.1 (t, C-7), 25.7 (t, C-6), 25.3 (t, C-1), 22.7 (t, C-12), 20.9 (q, C-20), 19.9 (t, C-2), 16.2 (q, C-17); *anal.* C 75.22%, H 9.24%, calcd for C<sub>20</sub>H<sub>30</sub>O<sub>3</sub>, C 75.43%, H 9.50%.

**Acknowledgment.** We thank Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) for financial assistance, and M.C. thanks CAPES/PICD for a fellowship. We also thank Dr. J. A. Maddry at TAACF for tuberculosis antimicrobial testing and Prof. L. H. B. Baptistella for helpful discussions.

## References and Notes

- (1) New address: M. Costa, Departamento de Química, UFRN, RN, Brazil.
- (2) Phadnis, A. P.; Patwardhan, S. A.; Dhaneshwar, N. N.; Tavale, S. S.; Row: T. N. G. *Phytochemistry* **1988**, *27*, 2899–2901.
- (3) Zhao, G.; Jung, J. H.; Smith, D. L.; Wood, K. V.; McLaughlin, J. L. *Planta Med.* **1991**, *57*, 380–383.
- (4) Chakrabarty, M.; Nath, A. C. *J. Nat. Prod.* **1992**, *55*, 256–258.
- (5) Hara, N.; Asaki, H.; Fujimoto, Y.; Gupta, Y. K.; Siongh, A. K.; Sahai, M. *Phytochemistry* **1995**, *38*, 189–194.
- (6) Rashid, M. A.; Hossain, M. A.; Hasan, C. M.; Reza, M. S. *Phytother. Res.* **1996**, *10*, 79–81.
- (7) Kijjoo, A.; Pinto, M. M. M.; Herz, W. *Planta Med.* **1989**, *55*, 205–206.
- (8) Kijjoo, A.; Pinto, M. M. M.; Pinho, P. M. M.; Tantisewie, B.; Herz, W. *Phytochemistry* **1990**, *29*, 653–655.
- (9) Kijjoo, A.; Pinto, M. M. M.; Pinho, M. M.; Herz, W. *Phytochemistry* **1993**, *34*, 457–460.
- (10) Hao, X.-J.; Lee, I.-S.; Chai, H.-B.; Farnsworth, N. R.; Soejarto, D. D.; Cordell, G. A.; Pezzuto, J. M.; Kinghorn, A. D. *Phytochemistry* **1994**, *37*, 1659–1662.
- (11) Hao, X.-J.; Yang, X.-S.; Zhang, Z.; Shang, L.-J. *Phytochemistry* **1995**, *39*, 447–448.
- (12) Bohlman, F.; Jakupovic, A. J.; King, R. M.; Robinson, H. *Rev. Latinoam. Quim.* **1984**, *15*, 16.
- (13) Habtemariam, S.; Gray, A. I.; Waterman, P. G. *Planta Med.* **1992**, *58*, 109–110.
- (14) Wijerathne, E. M. K.; da Silva, L. B.; Tezuka, Y.; Kikuchi, T. *Phytochemistry* **1995**, *39*, 443–445.
- (15) Costa, M.; Tanaka, C. M. A.; Imamura, P. M.; Marsaioli, A. J. *Phytochemistry* **1999**, *50*, 117–122.
- (16) Hagikawa, H.; Inome, K.; Uda, H. *J. Chem. Soc., Perkin Trans. 1* **1995**, 757–764.
- (17) For a similar synthesis of a sesterterpene hydroxybutenolide see: Piers, E.; Wai, J. S. M. *Can. J. Chem.* **1994**, *72*, 146–157.
- (18) Pantarotto, H.; Imamura, P. M. *Liebigs Ann.* **1995**, 1891–1894.
- (19) Nunes, F. M. N.; Imamura, P. M. *J. Braz. Chem. Soc.* **1996**, *7*, 181–186.
- (20) Santos, C.; Rosso, C. R. S.; Imamura, P. M. *Synth. Commun.* **1999**, *29*, 1903–1910.
- (21) Since **2a** was the starting material, the configurations of the synthetic compounds **5** and **6** are enantiomeric to those of the natural hydroxybutenolides.
- (22) Costa, M.; Fujiwara, F. Y.; Imamura, P. M. *Magn. Reson. Chem.* **1998**, *36*, 542–544.
- (23) Kernan, M. R.; Faulkner, D. J. *J. Org. Chem.* **1988**, *53*, 2773–2776.
- (24) Sarma, D. N.; Sharma, R. P. *Tetrahedron Lett.* **1985**, *26*, 2581–2584.
- (25) Walker, R. P.; Faulkner, D. J. *J. Org. Chem.* **1981**, *46*, 1098–1102.
- (26) Rosser, R. M.; Faulkner, D. J.; Bass, L. S.; Cun-Heng, H.; Clardy, J. *J. Org. Chem.* **1984**, *49*, 5160–5163.
- (27) Tuberculosis antimicrobial testing was performed at TAACF, Southern Research Institute, Birmingham, AL.

NP000105F